

Self-promoted and self-terminated PIN2-mediated auxin transport is crucial during coordination of pavement cell (PC) morphogenesis

Patricio Pérez-Henríquez^{1,2}, Wenxin Tang^{1,2}, Shingo Nagawa², Lucia Strader³, Zhenbiao Yang^{1,2}

¹ Center for Plant Cell Biology, Institute of Integrated Genome Biology, and Department of Botany and Plant Sciences, University of California, Riverside, CA 92521, USA.

² FAFU-UCR Joint Center for Horticultural Biology and Metabolomics, Haixia Institute of Science and Technology, Fujian Agriculture and Forestry University, Fuzhou, Fujian, China

³ NSF Science and Technology Center for Engineering Mechanobiology, Department of Biology, Washington University in St. Louis, St. Louis, MO 63130, USA

INTRODUCTION

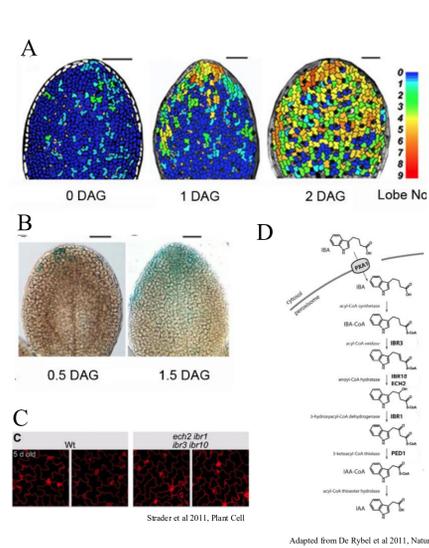


Figure 1. Progressive tip-to-base activation of pavement cell (PC) interdigitation is preceded by tip-high auxin transcriptional response.

(A) Progression of pavement cell morphogenesis (interdigitation) occurs gradually over time within 48 h (B) Auxin transcriptional response showed highest level before initiation of PC morphogenesis (C) IBA to IAA conversion guided by enoyl-coA hydratase enzymes and paralogues is crucial for PC morphogenesis (D) IBA to IAA conversion pathway. Notice key enzyme enoyl-coA hydratase, ECH2. The conversion occurs within peroxisomes.

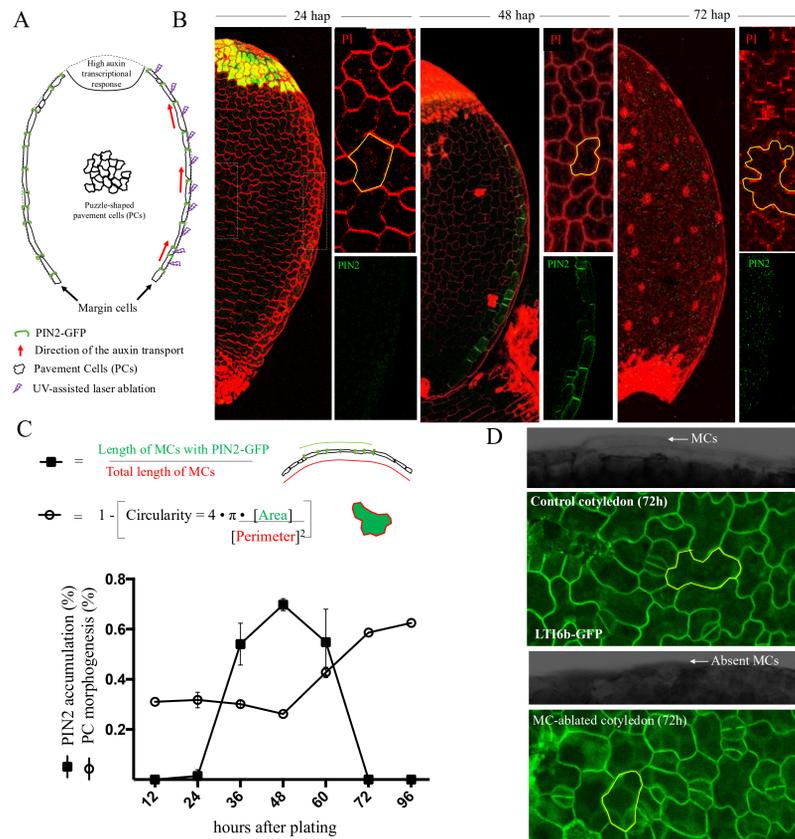


Figure 2. PIN2 accumulation in MCs precedes PC morphogenesis and its function is crucial during global coordination of pavement cell morphogenesis

(A) Scheme of cotyledon. (B) Time lapse identifying PIN2-GFP accumulation along the MCs and the progression of pavement cell morphogenesis (C) Quantification of PIN2 accumulation along cotyledon's margin cells (MCs) (D) Pavement cells when margin cell are laser-assisted ablated (E) Circularity of pavement cells at the tip of the cotyledon of *pin2* mutants (F) Quantification of images in E.

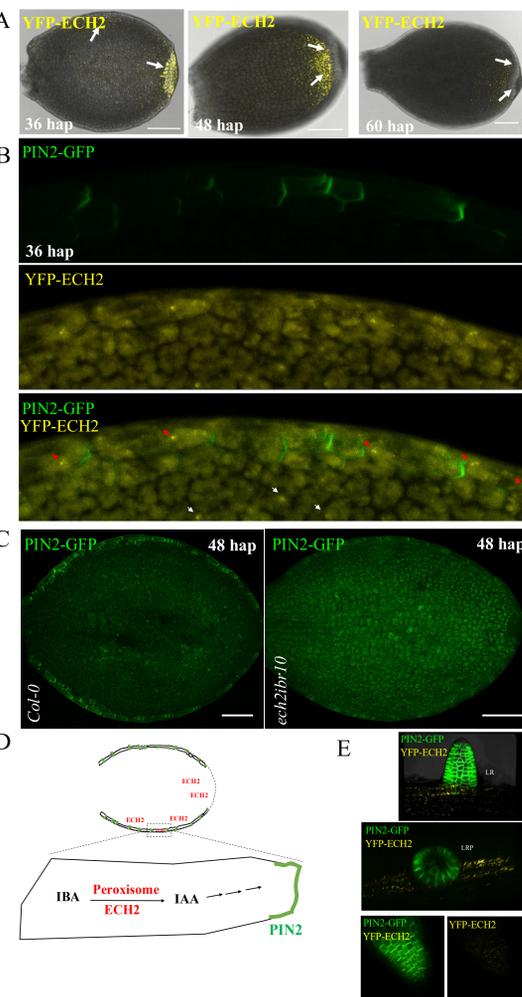


Figure 4. IBA-derived auxin regulates PIN2 accumulation along cotyledon's margin cells.

(A) YFP-ECH2 accumulation in the borders and tip of expanding cotyledon. (B) YFP-ECH2 accumulates at MCs suggesting IBA→IAA conversion leading to PIN2-GFP accumulation (C) IBA-to-IAA conversion double mutant *ech2 ibr10* showed diminished PIN2-GFP levels at MCs (D) Scheme for relationship between IBA/IAA and PIN2 levels (E) ECH2 accumulates in other developing /expanding organs

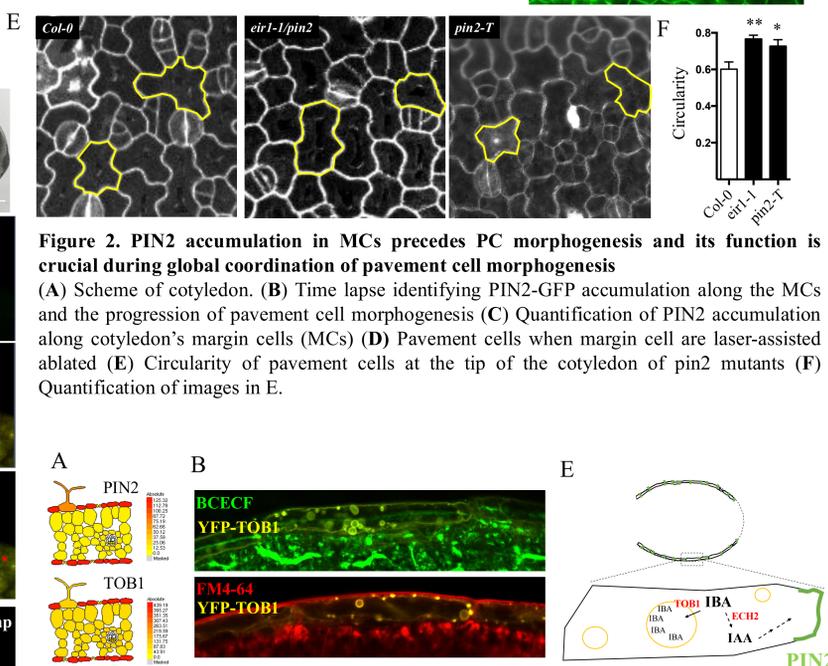
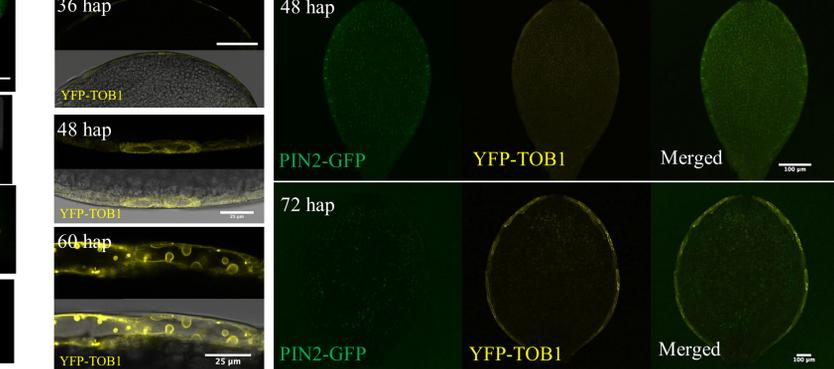


Figure 5. TOB1-mediated vacuole internalization of IBA terminates with PIN2 polar accumulation at cotyledon's margin cells.



(A) Expression levels for IBA transporter TOB1 is high in epidermis, similar to PIN2 (data from <http://efp.ucr.edu/>). (B) YFP-TOB1 accumulates in vacuoles within MCs according to tonoplast marker FM464 and vacuolar lumen marker BCECF (C) YFP-TOB1 consolidates vacuolar localization towards 60 hap (D) YFP-TOB1 and PIN2-GFP accumulation temporal patterns correlates inversely. Together this evidence suggest that vacuolar TOB1 contributes to reduce available cytoplasmic IBA which reduces PIN2-levels

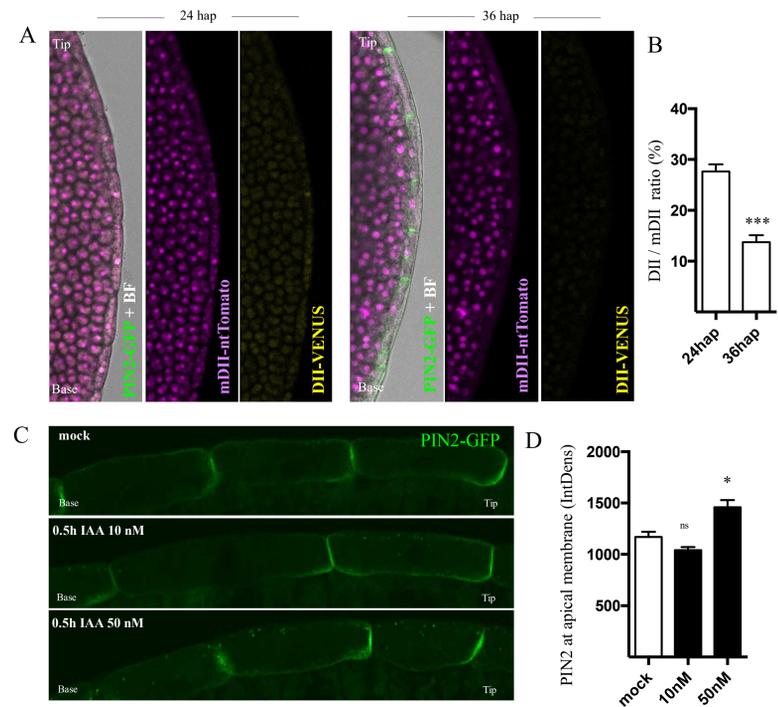


Figure 3. Auxin promotes its own PIN2-mediated efflux along cotyledon's margin cells.

(A) Auxin signaling reporter indicate increased auxin signaling after PIN2-GFP accumulation (B) Quantification of ratio metric R2D2 signal in the margin cells (C) PIN2-GFP levels at the apical membrane after 30 min of exogenously applied auxin IAA (D) Quantification of PIN2 at the apical membrane of MC cells.

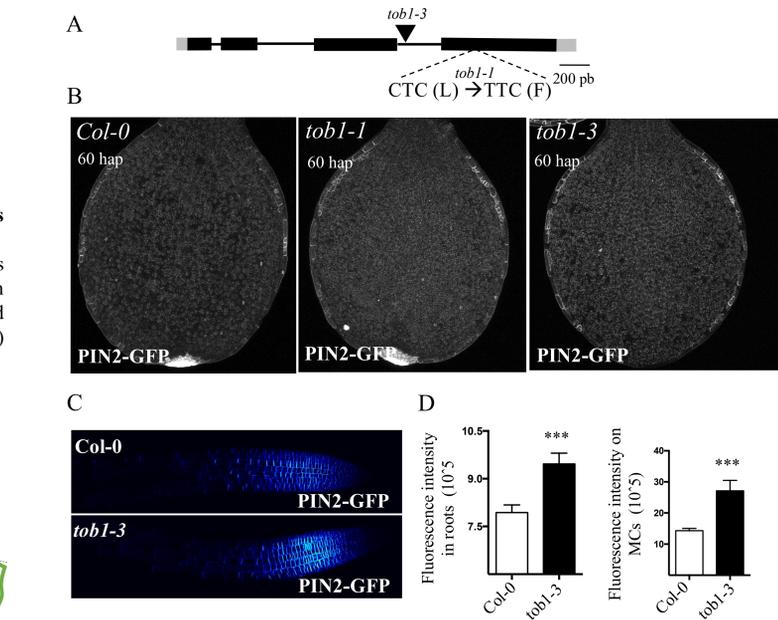
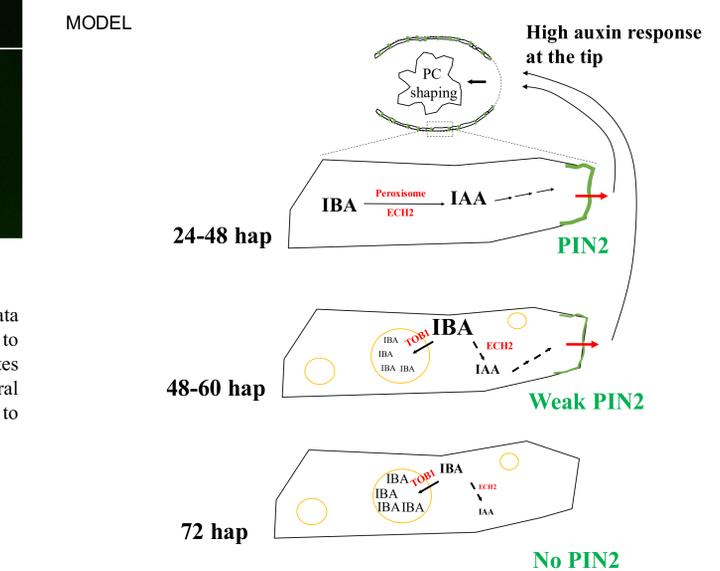


Figure 6. TOB1 function to downregulate PIN2 levels.

(A) Gene model for TOB1 depicting *tob1-1* point mutation and *tob1-3* insertional mutation (B) loss-of-function mutants *tob1-1* and *tob1-3* cause increased PIN2-GFP accumulation on MCs (C) *tob1-3* showed increased PIN2-accumulation at root tips (D) Quantification of PIN2 level in Col-0 and *tob1-3* background.



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REFERENCES

Li *et al.* in preparation (Yang Lab)
Strader *et al.* 2011. Plant Cell
De Rybel *et al.* 2011. Nature Chem Biol

CONCLUSION

1. Auxin transporter PIN2 accumulates transiently at cotyledon's margin cells preceding PC morphogenesis
2. PIN2 functions to mobilize auxin self-promoting its own transport towards the tip of the cotyledon and its function is critical for PC morphogenesis.
3. IBA-derived auxin, through ECH2 enzyme, is critical in determining PIN2 levels and PC morphogenesis
4. TOB1 is an IBA transporter that reduces cytoplasmic IBA after internalizing IBA into vacuoles.